

8° WORKSHOP IN EMATOLOGIA TRASLAZIONALE

DELLA SOCIETÀ ITALIANA DI EMATOLOGIA SPERIMENTALE

Firenze - Auditorium CTO - A.O.U. Careggi, 22-23 giugno 2023

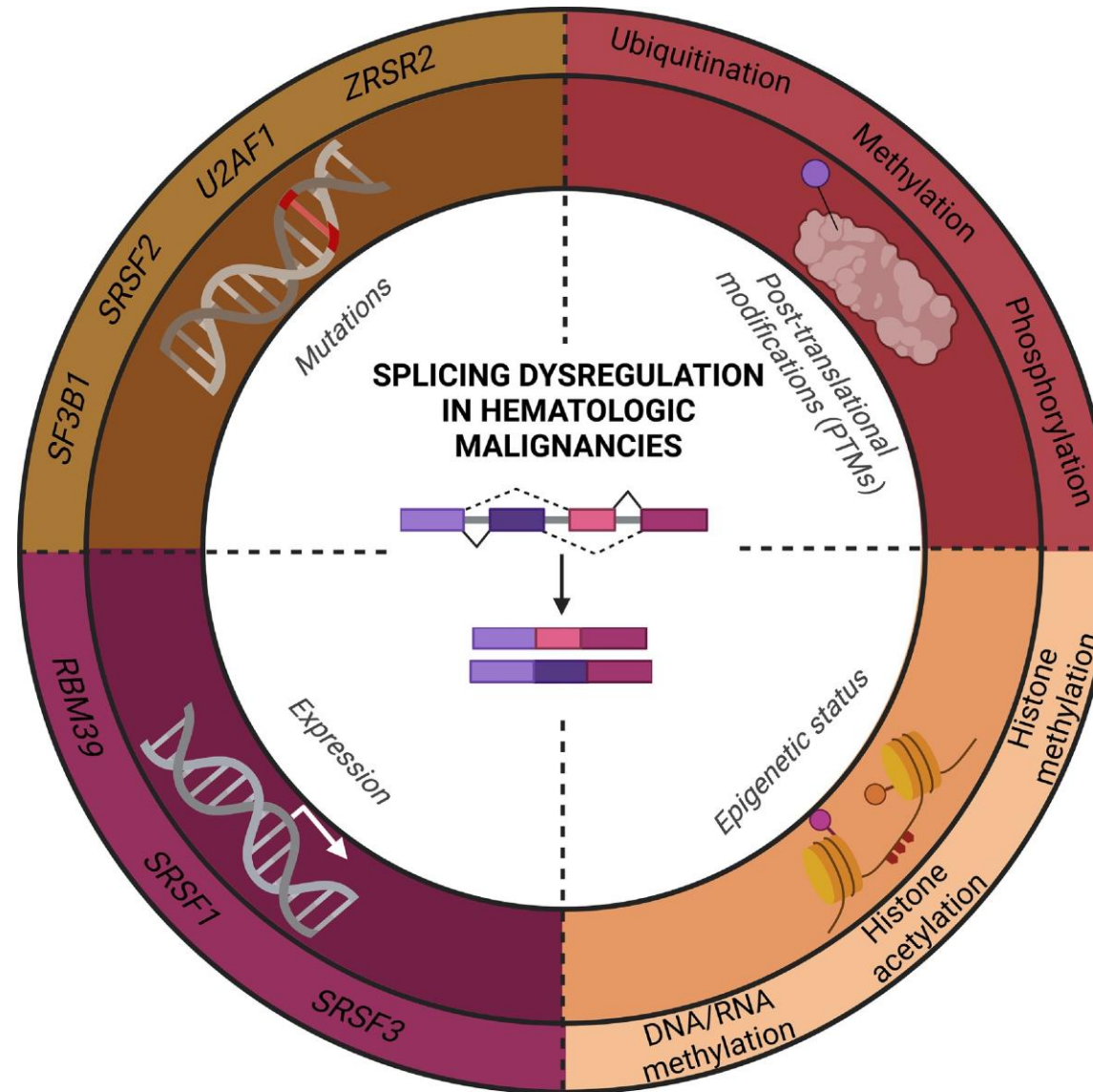


Alterazioni dello Splicing nelle MDS a basso rischio

Emiliano Fabiani, PhD

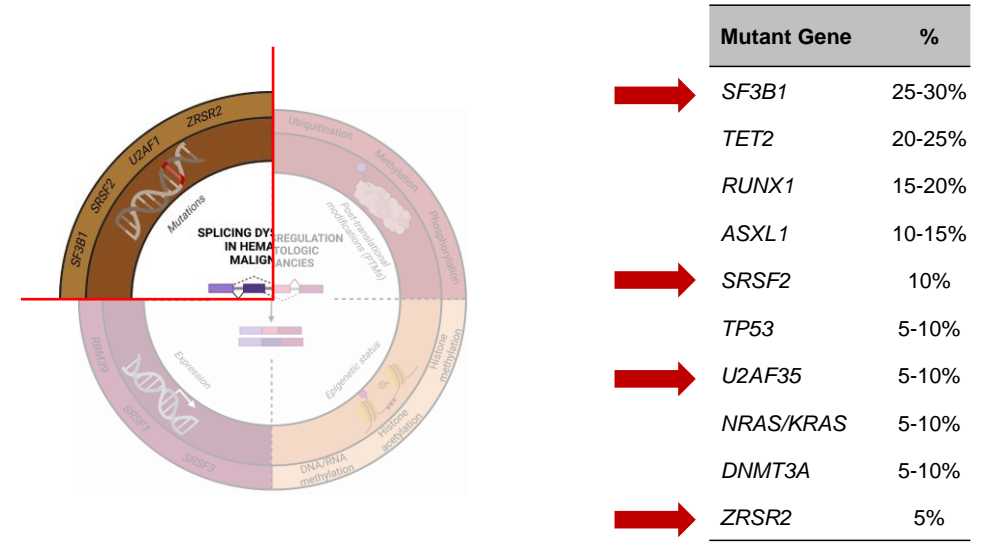
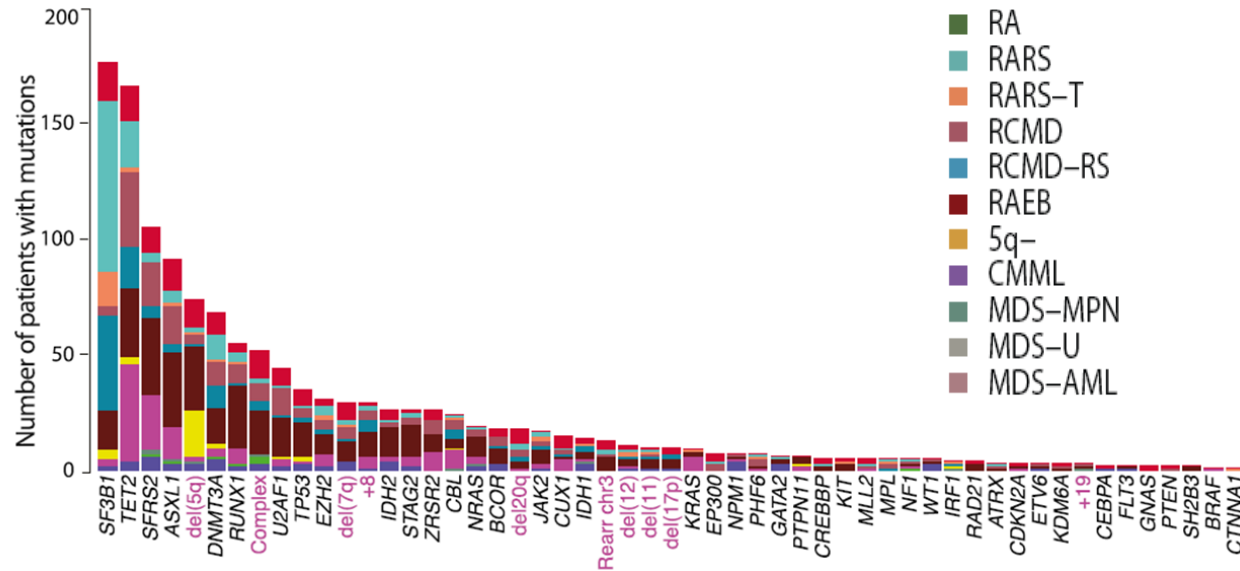


Molecular mechanisms that can drive aberrant splicing in hematologic malignancies

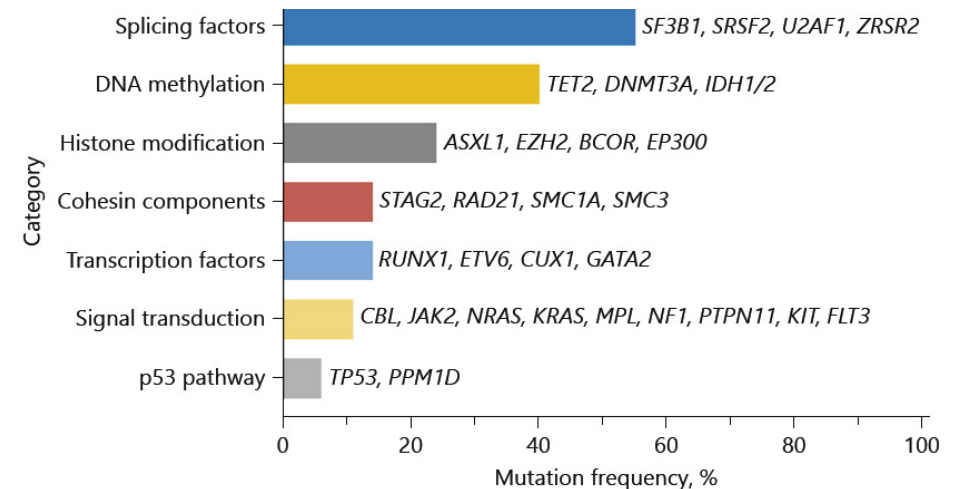




Mutational landscape of MDS

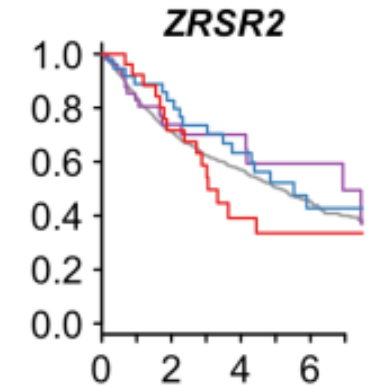
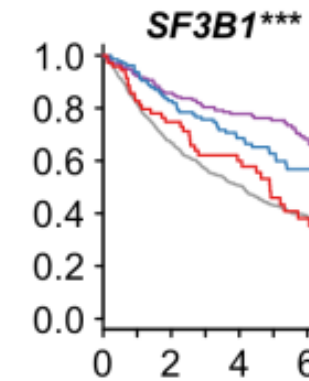
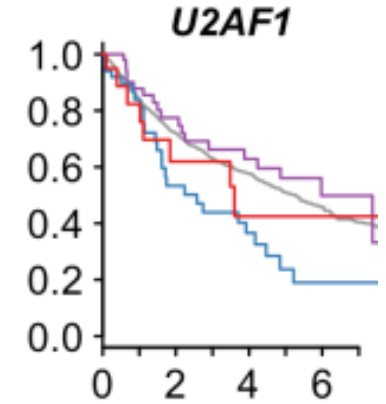
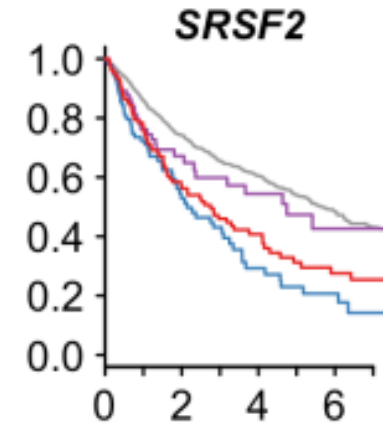
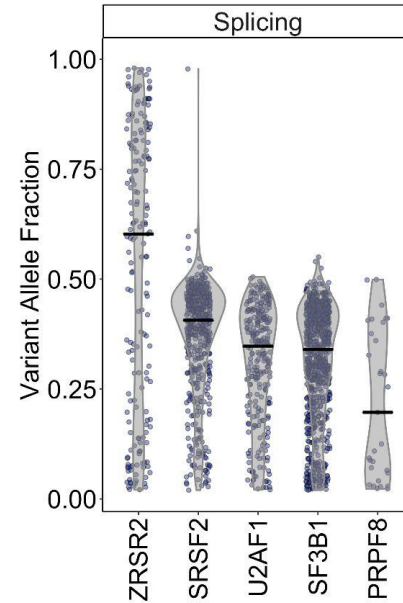
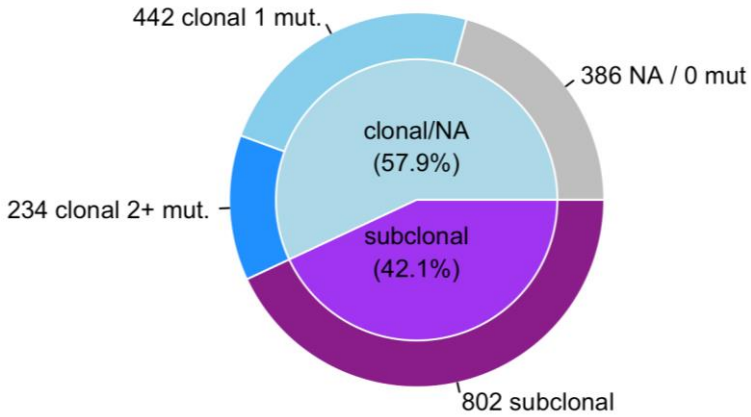


- About 95% of patients with MDS have at least one mutation (t-NGS 30-80 genes)
- Genes belonging to the splicing machinery (SF3B1, SRSF2, U2AF1 and ZRSR2) are the most frequently mutated genes in MDS (50-60%)
- SF3B1 is the most frequently mutated gene in MDS (25-35%)





Survival between clonal and subclonal mutations



—	wt	.	P (0.05, 0.1]
—	clonal	*	P (0.01, 0.05]
—	indetermined	**	P (0.001, 0.01]
—	subclonal	***	P < 0.001

- No significant difference in survival between clonal and subclonal mutations for SRSF2, U2AF1 and ZRSR2
- The worse survival associated with subclonal SF3B1 suggests it belongs to a separate bystander clone, with a main clone driven by other mutations

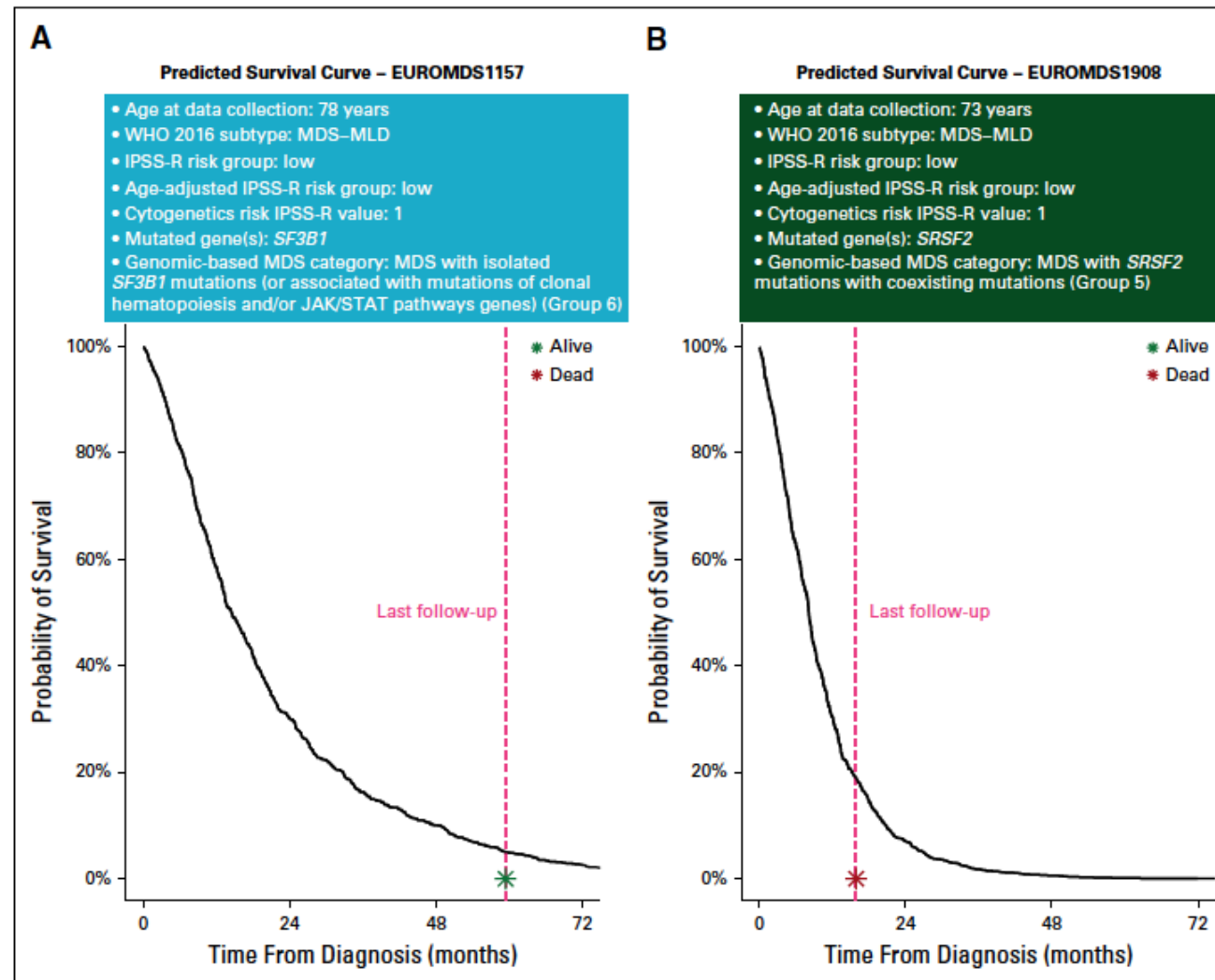


Impact of specific splicing mutations in LR-MDS



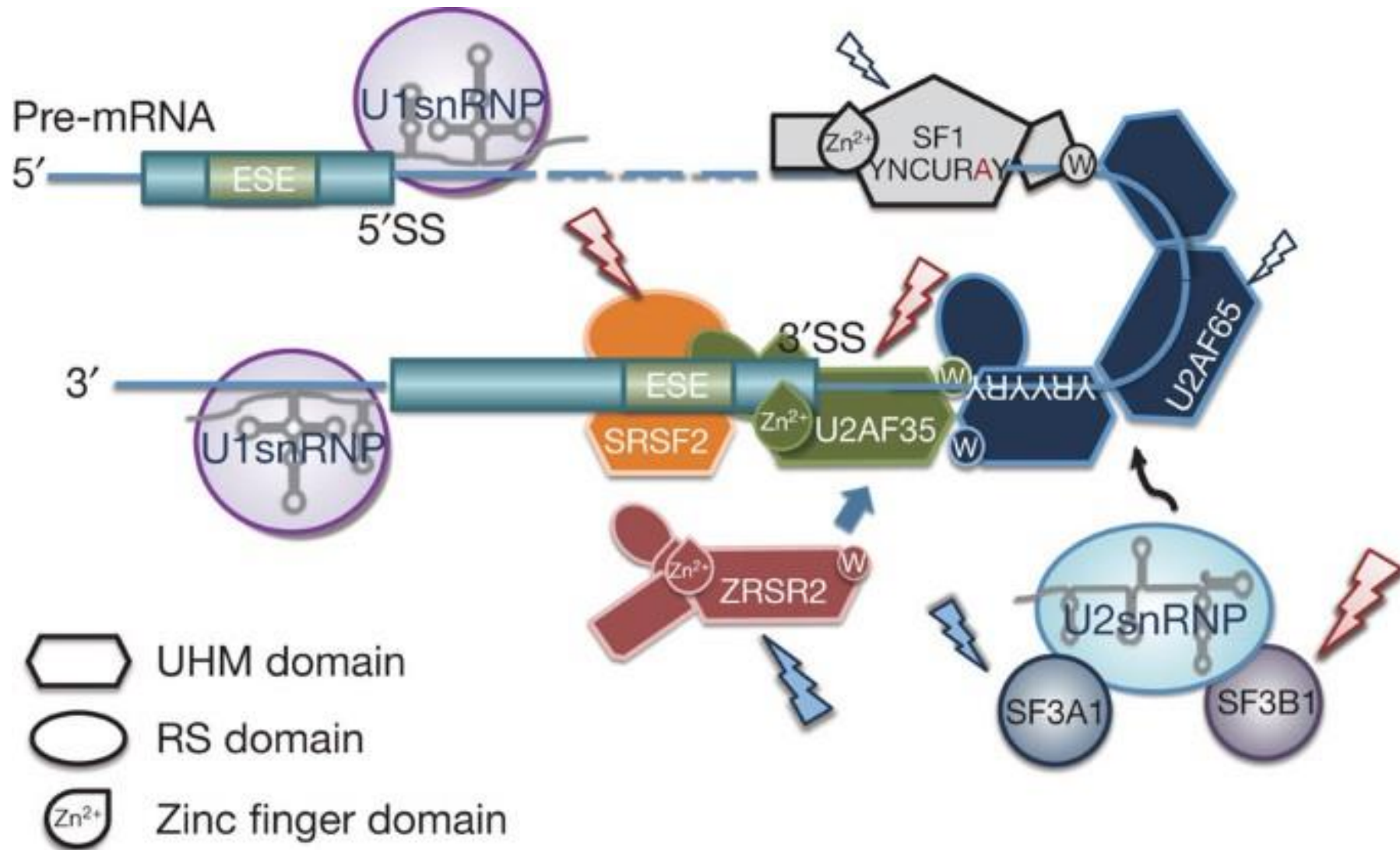
SF3B1^{mut} Group 6

SRSF2^{mut} Group 5



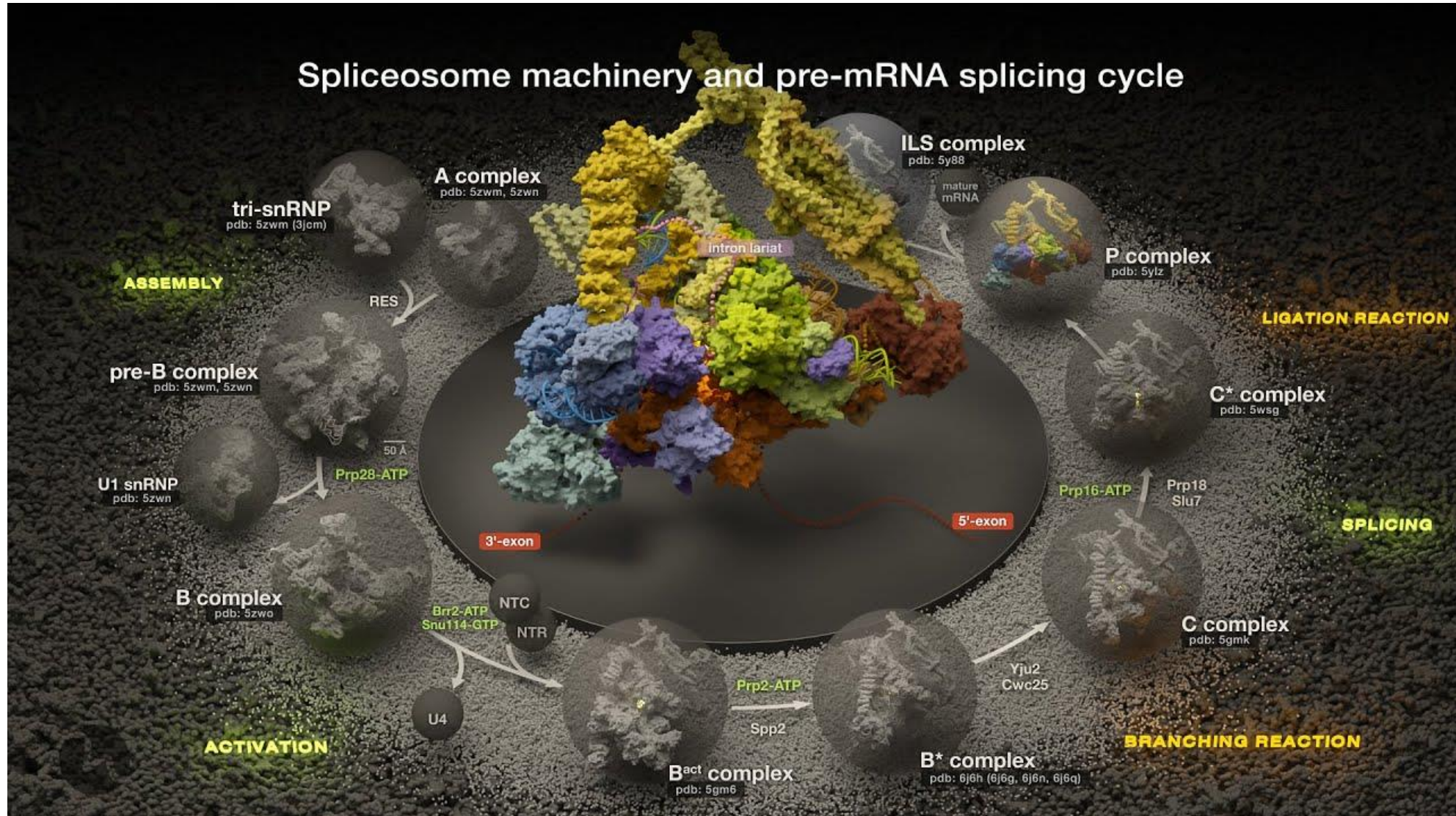


Splicing machinery



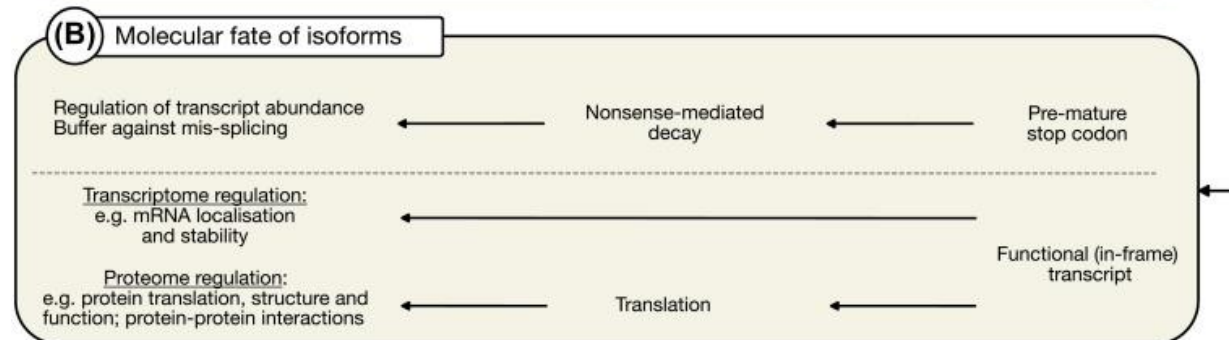
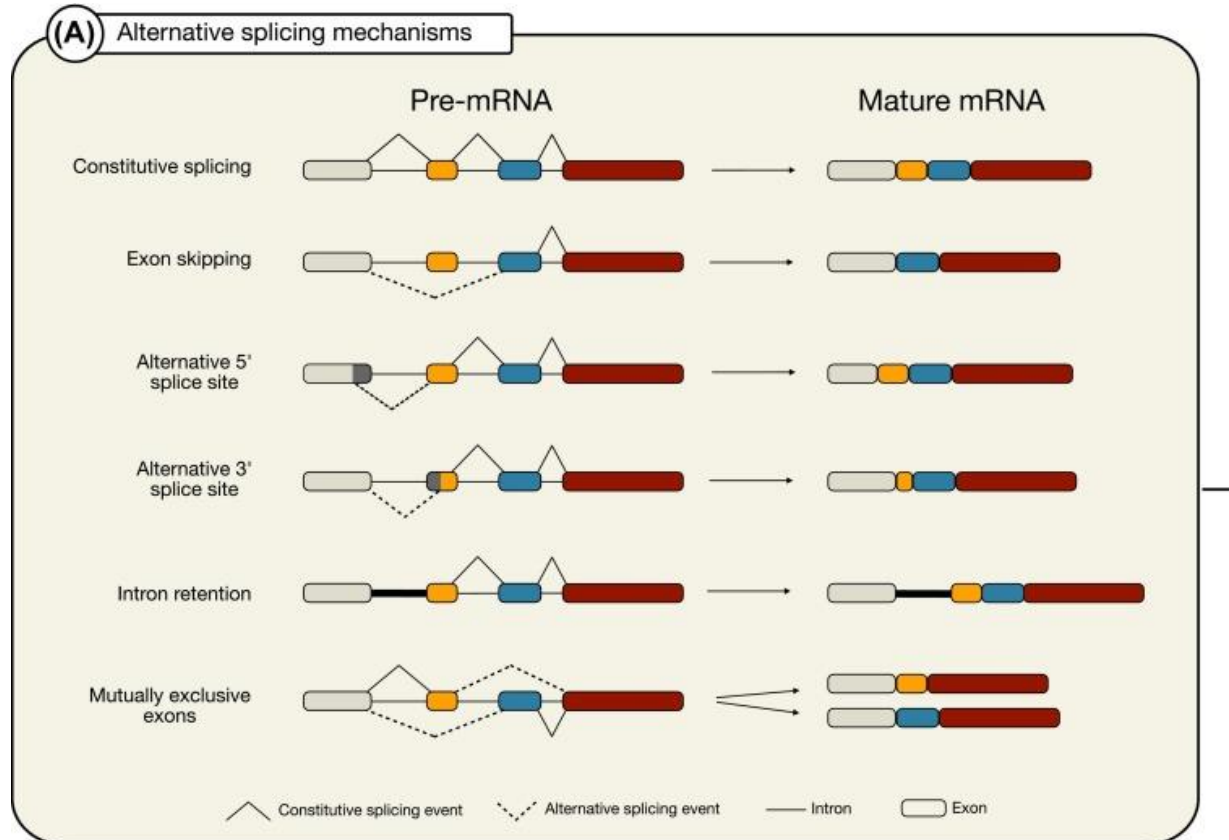


Splicing machinery



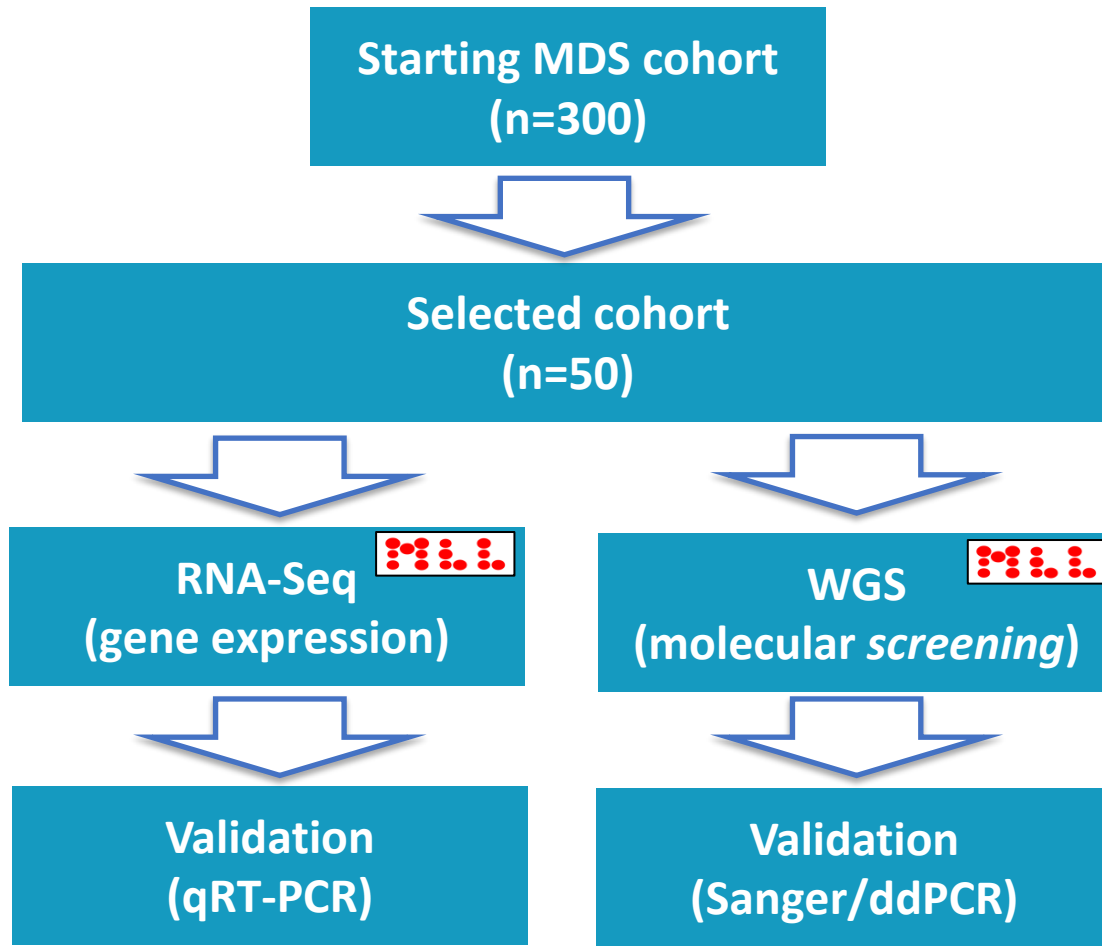


Alternative splicing events





Study cohort



Patients' selection according to IPSS-R and splicing factors mutation profile

Gene	Target region (exon)	Gene	Target region (exon)	Gene	Target region (exon)
ABL	4-9	FLT3	13-15 and 20	PTPN11	3,7-13
ASXL1	9,11,12	HRAS	2,3	RUNX1	all
BRAF	15	IDH1	4	SETBP1	4
CALR	9	IDH2	4	SF3B1	10-16
CBL	8,9	JAK2	all	SRSF2	1
CEBPA	all	KIT	2,8-11, 13,17 and 18	TET2	all
CSF3R	all	KRAS	2,3	TP53	all
DNMT3A	all	MPL	10	U2AF1	2,6
ETV6	all	NPM1	10,11	WT1	6-10
EZH2	all	NRAS	2,3	ZRSR2	all



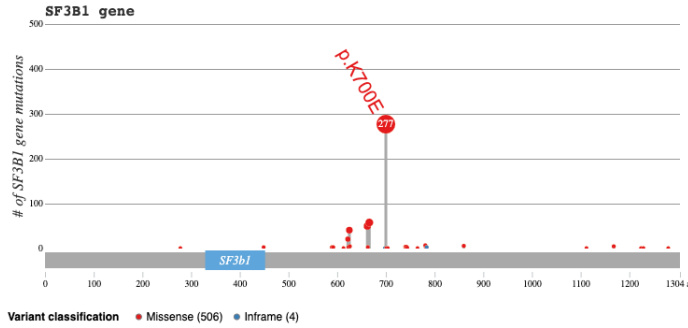
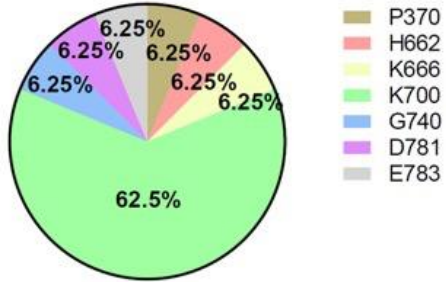
Coverage	1000X
*VAF	> 1%



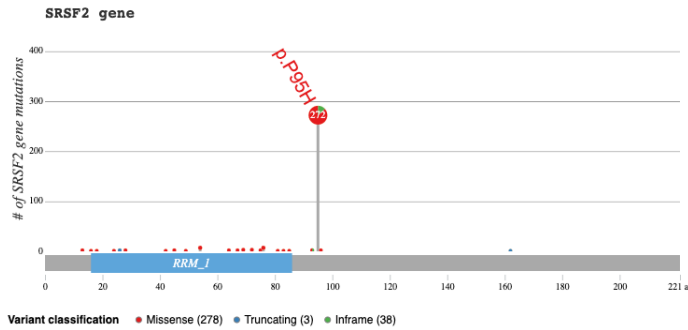
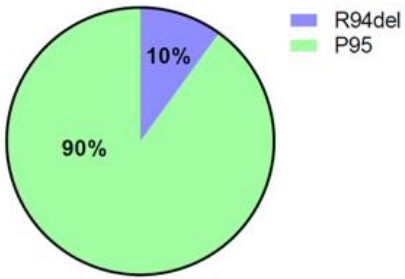
WGS analysis: mutation types and co-mutations pattern



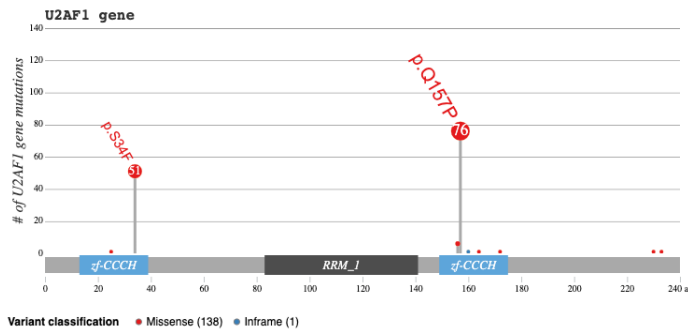
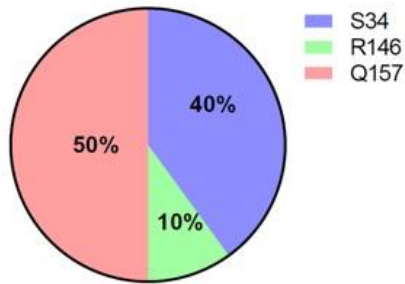
Type of mutations in SF3B1 gene



Type of mutations in SRSF2 gene

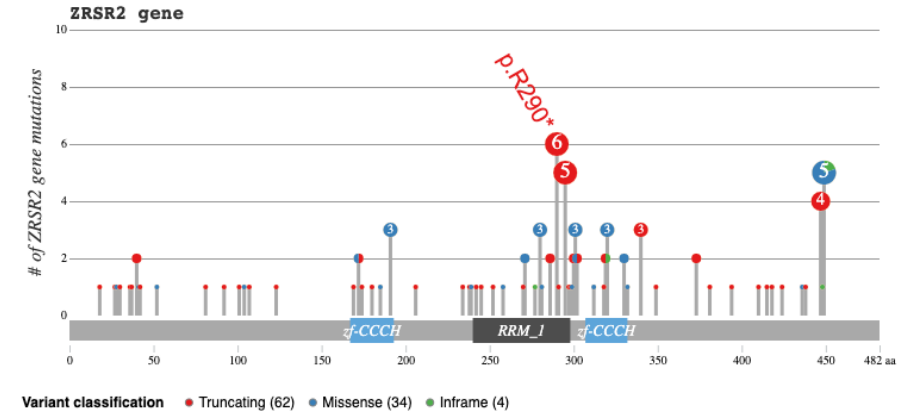


Type of mutations in U2AF1 gene



Exclusion criteria

ZRSR2 mutated patients



SF co-mutated patients

Patient ID	Mutations	VAF (%)	CO-Mutations	VAF (%)
UPN2	SF3B1 E738K	33,6	SRSF2 P95H	21
UPN13	SF3B1 P370T	4,5	SRSF2 P95H	31,4
UP16	U2AF1 Q157P	39,1	ZRSR2 K405Rfs*	74,2
UPN17	U2AF1 Q157P	11,7	ZRSR2 W340*	30
UPN27	SF3B1 K700E	42,1	SRSF2 R94H100del	29,9

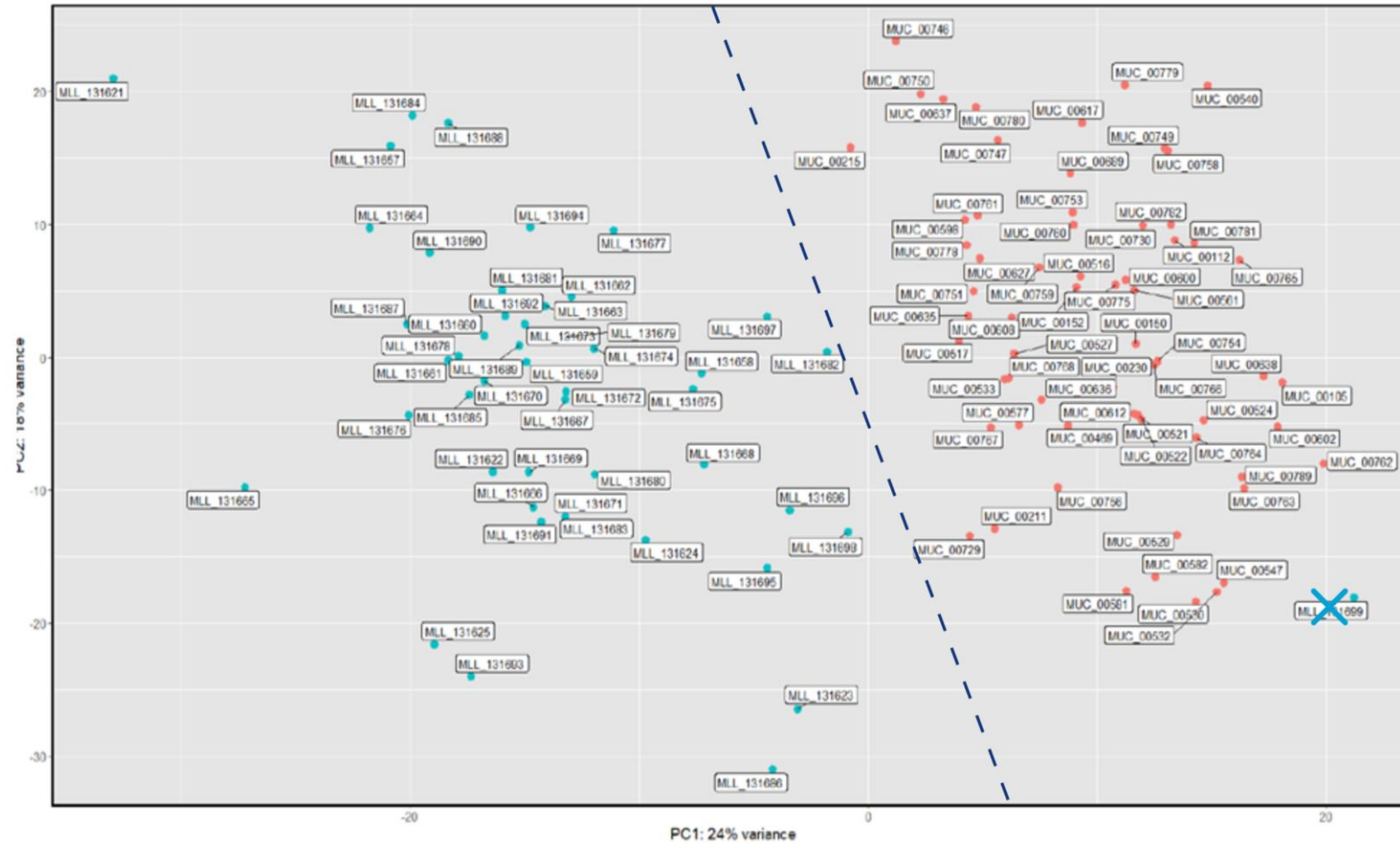


Differentially expressed genes by RNA-Seq analysis



MDS patients

Controls



$p\text{-adj} < 0,05; \text{Log}_2 \text{FC} \geq 2 \text{ e } \text{log}_2 \text{FC} \leq -2$

LR-MDS vs CTRL

Down
3921

UP
300

SF3B1 mut vs SF WT

Down
10

UP
612

SRSF2 mut vs SF WT

Down
6

UP
7

U2AF1 mut vs SF WT

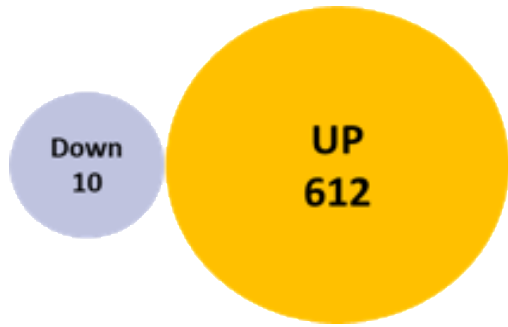
UP
4



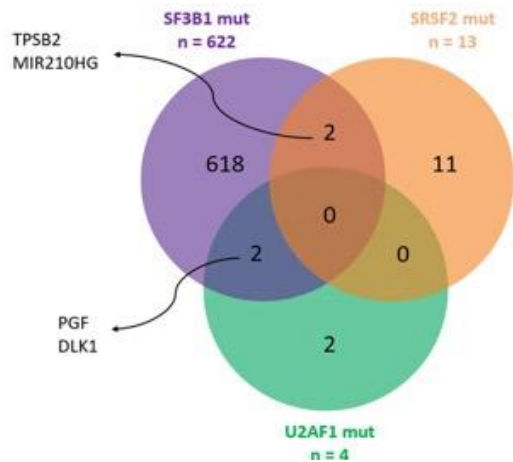
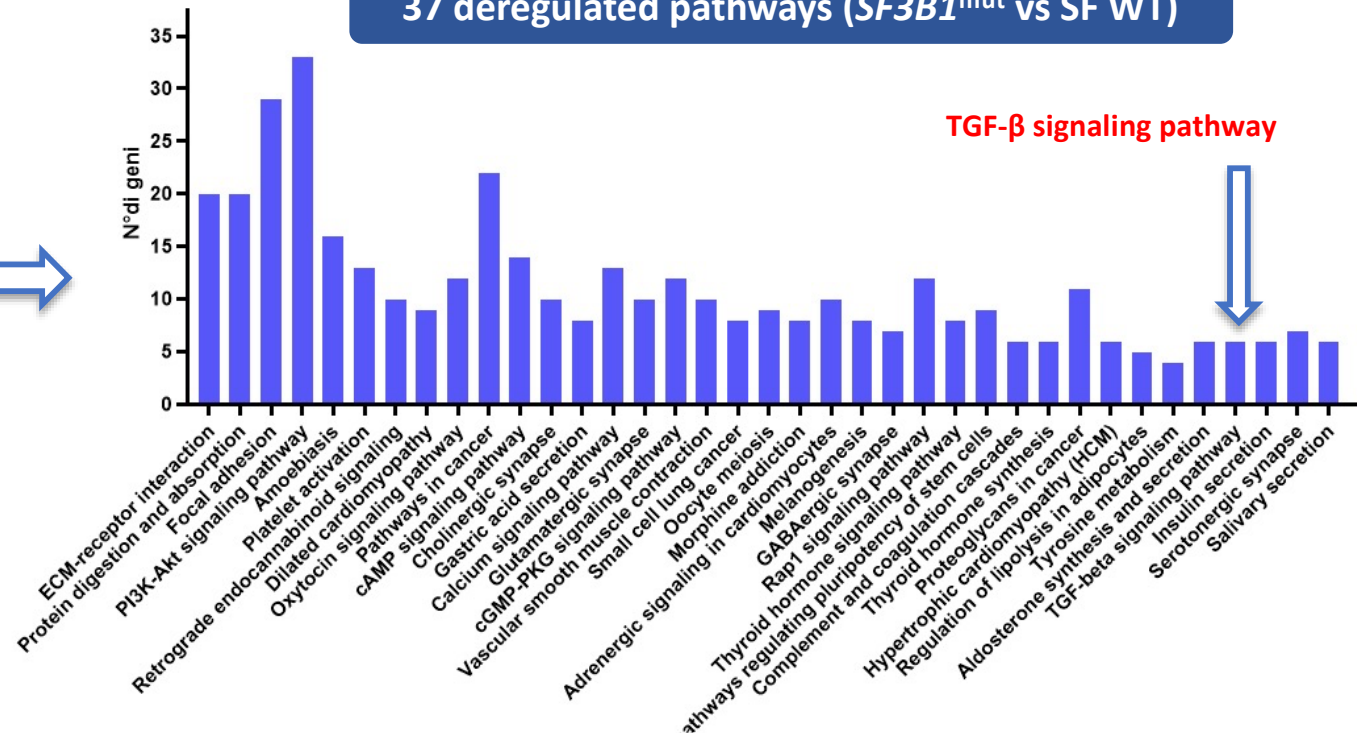
Differentially expressed genes in LR-MDS SF3B1^{mut}



SF3B1^{mut} (13) vs SF WT (8)

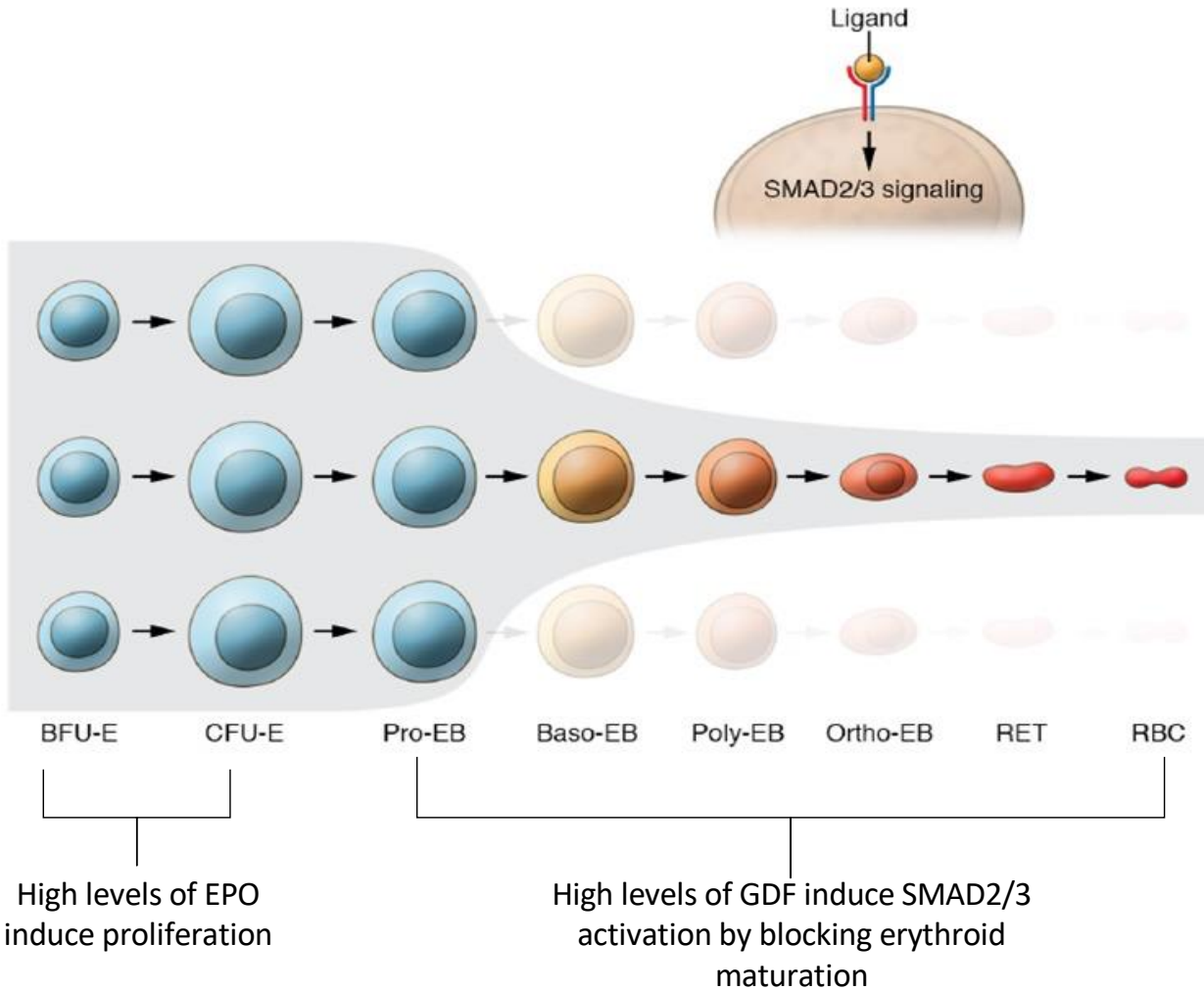


37 deregulated pathways (SF3B1^{mut} vs SF WT)

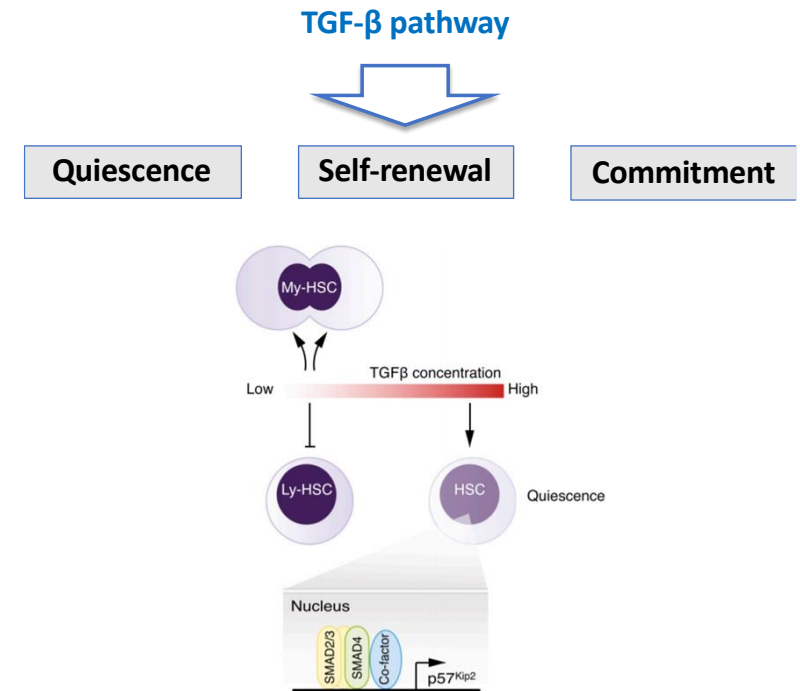




TGF- β pathway and modulators in SF3B1^{mut}

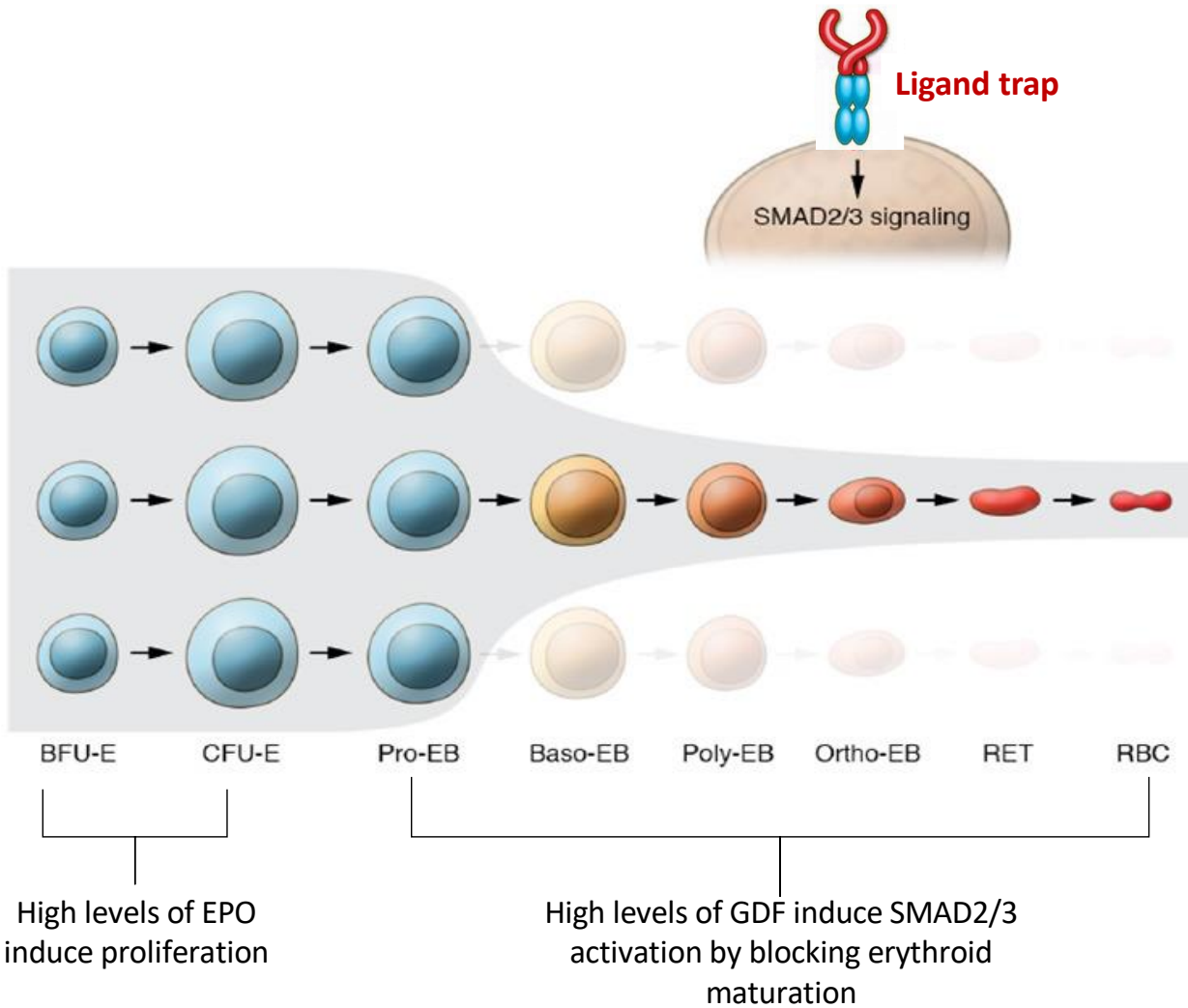


- Transforming growth factor beta (TGF- β) signaling pathway is key to hematopoiesis regulation
- Up-regulation of TGF- β signaling has been proposed as one of the causes of ineffective hematopoiesis



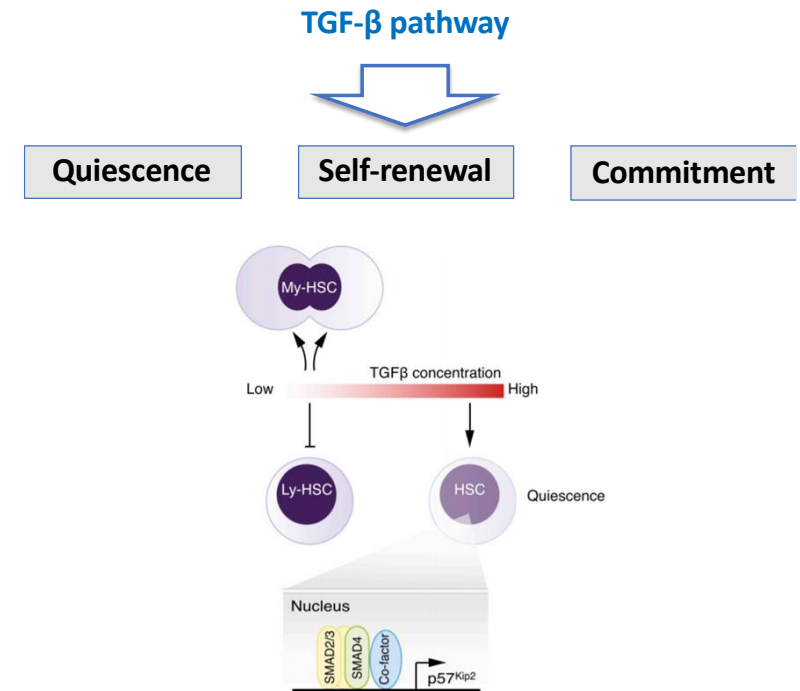


TGF- β pathway and modulators in SF3B1^{mut}



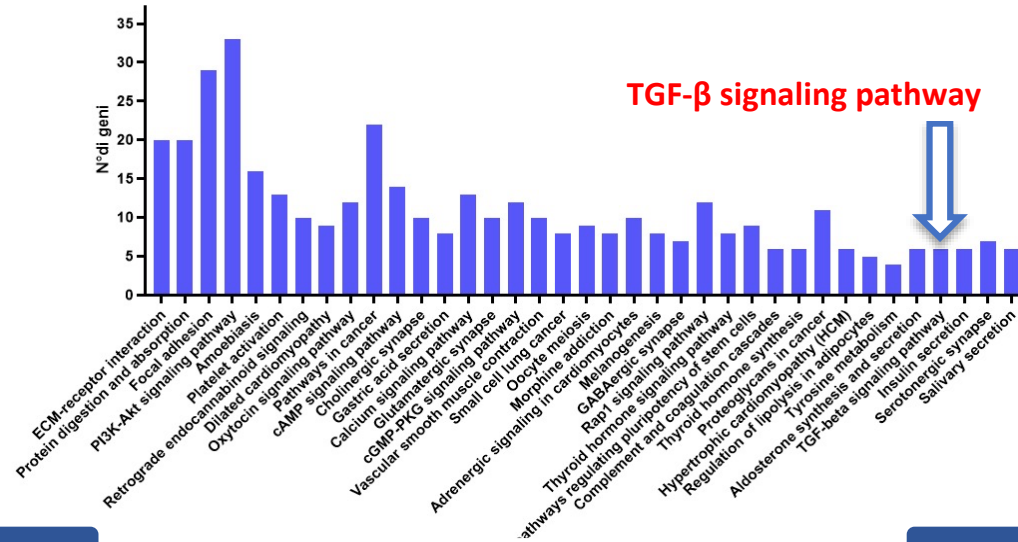
Ligand trap

- It binds to GDF11 and other members of the TGF- β superfamily, inhibiting their binding to the activin IIB receptor.
- Thus, it prevents the signal activation of SMAD2 and SMAD3.

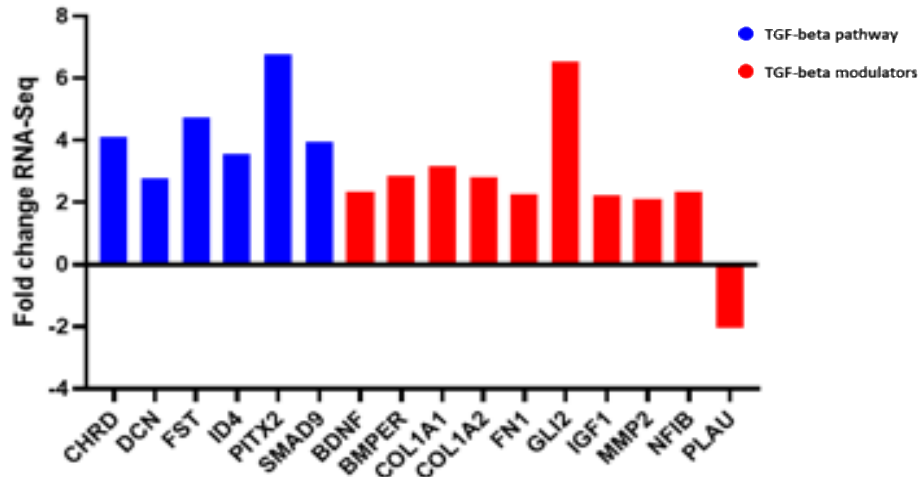




TGF- β pathway and modulators in SF3B1^{mut}



TGF-beta pathway and modulators



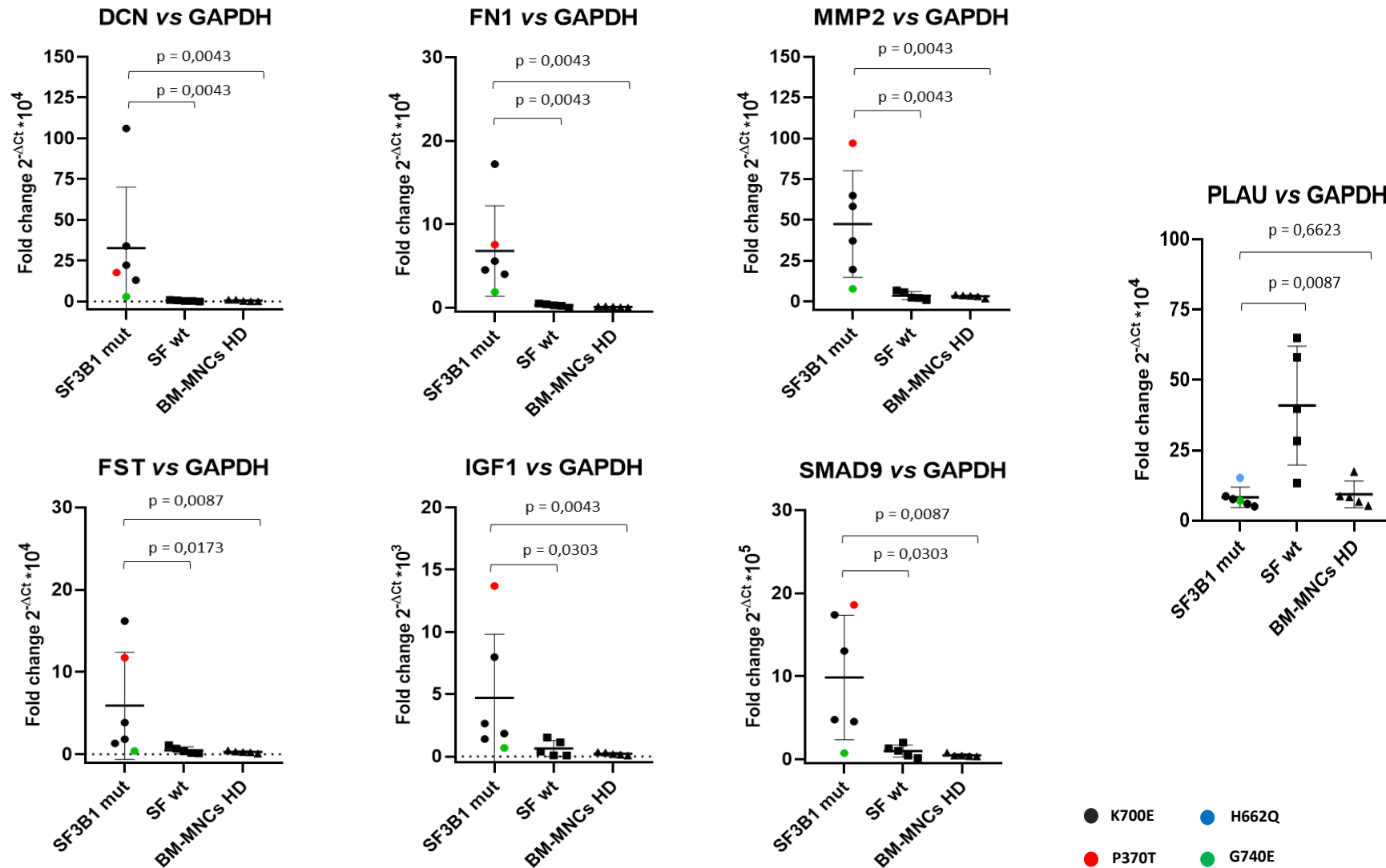
Validation

Q-RT-PCR validation

- SMAD9** (SMAD family member 9)
- DCN** (Decorin)
- FST** (Follistatin)
- FN1** (Fibronectin)
- IGF1** (Insulin-like growth factor I)
- MMP2** (Matrix metalloproteinase 2)
- PLAU** (Plasminogen activator, urokinase)

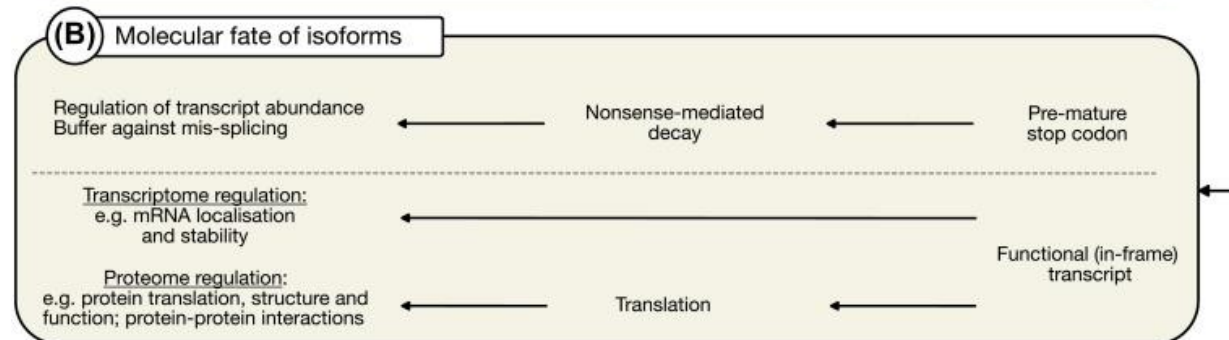
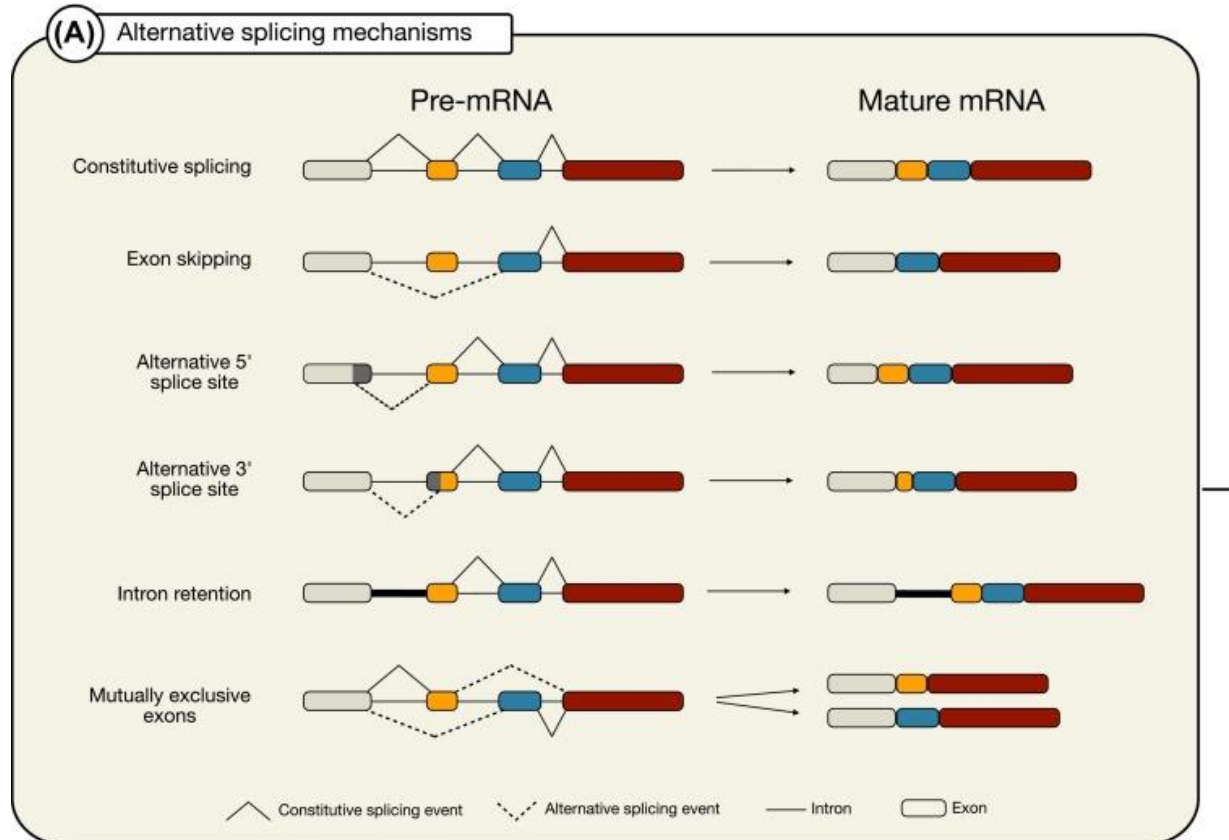


TGF- β pathway and modulators in SF3B1^{mut}





Alternative splicing events

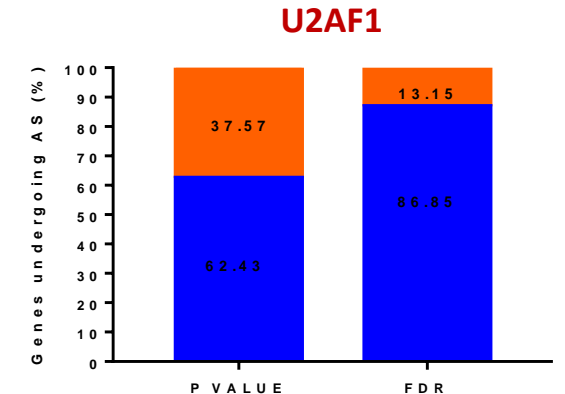
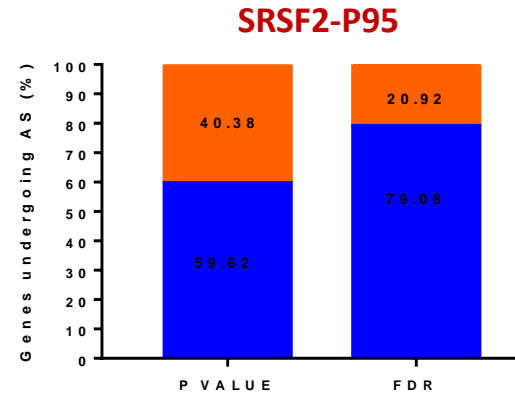
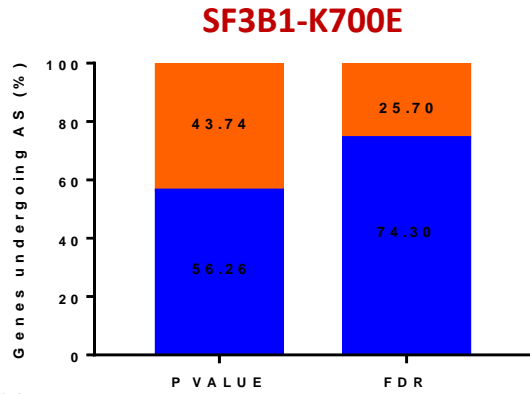




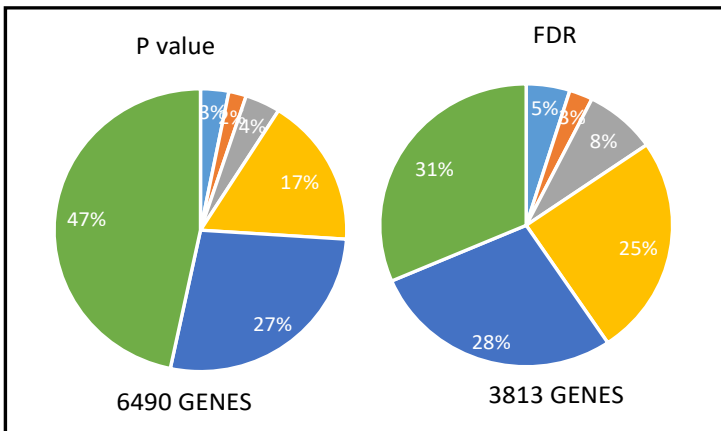
Percentage of genes subjected to AS regulation

■ Regulated
■ Unregulated

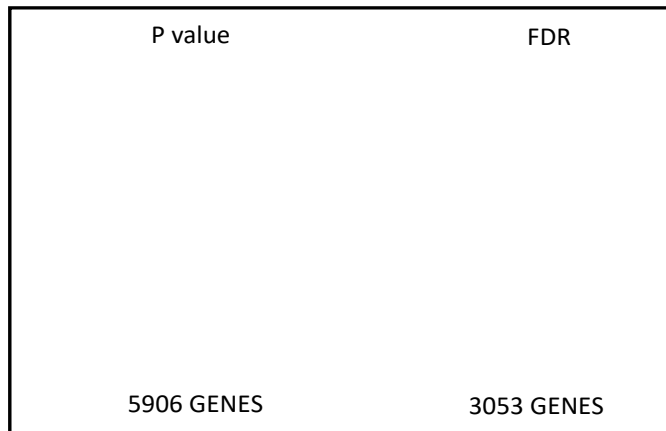
A3SS: alternative 3'ss
A5SS: alternative 5'ss
RI: intron retention
SE: exon skipping
MXE: mutually exclusive exons
EV MIX: >1 AS events



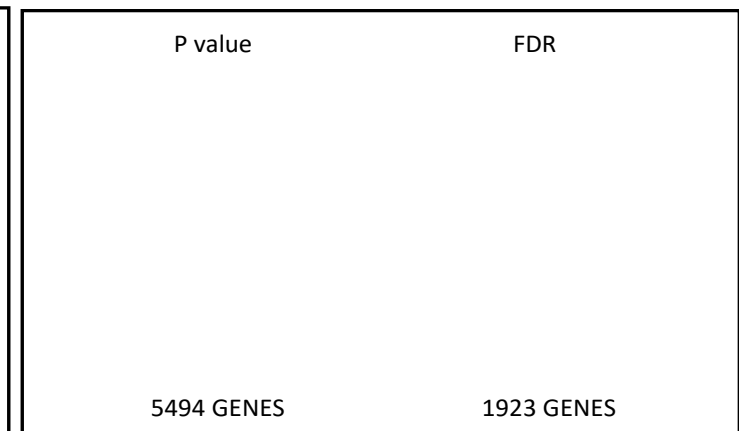
SF3B1-K700E MDS



SRSF2 MDS



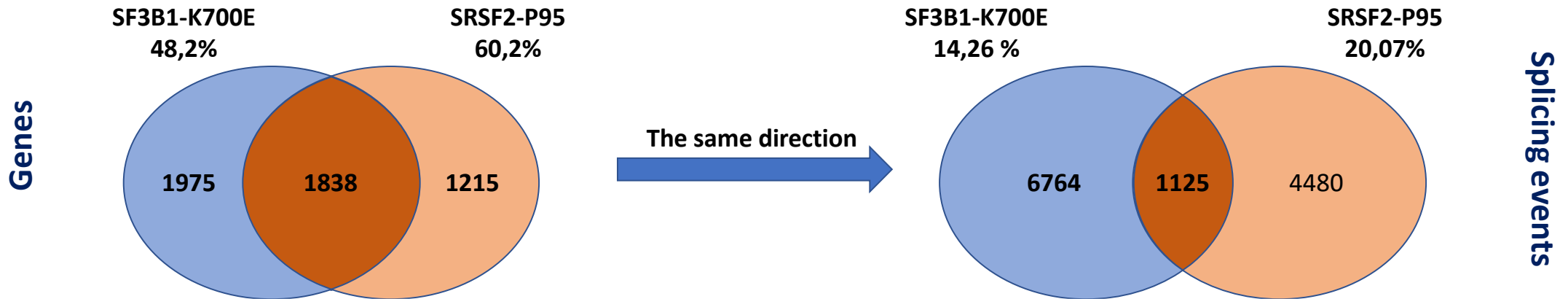
U2AF1 MDS



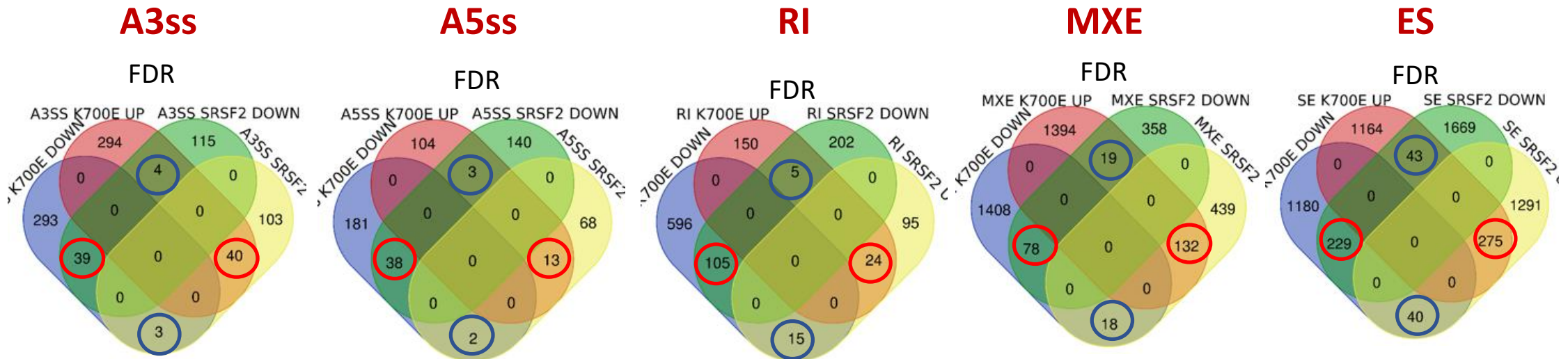
A3SS A5SS RI MXE SE EV MIX



Overlap of genes/events undergoing AS regulation in SF3B1-K700E vs SRSF2-P95 mutated patients



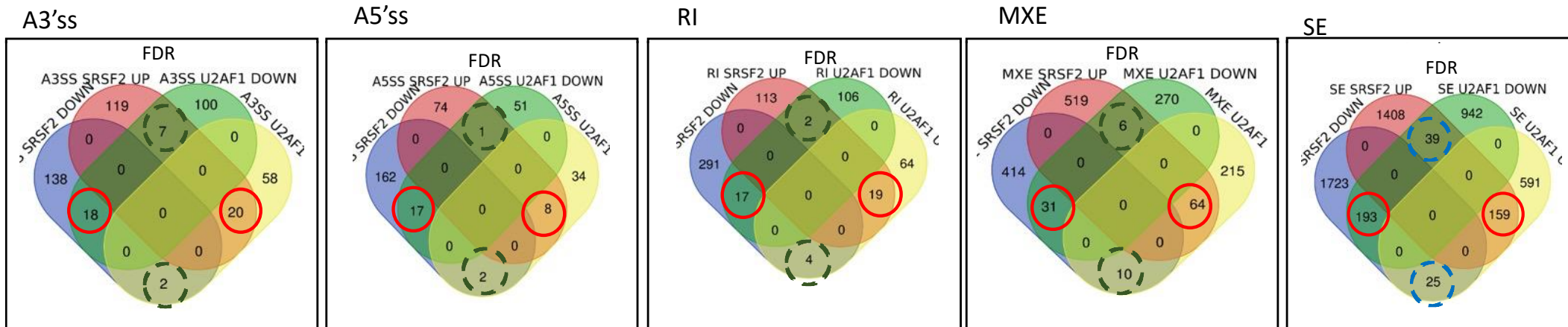
Red dashed circle = AS regulated in the same direction; Blue dashed circle = AS regulated in opposite direction



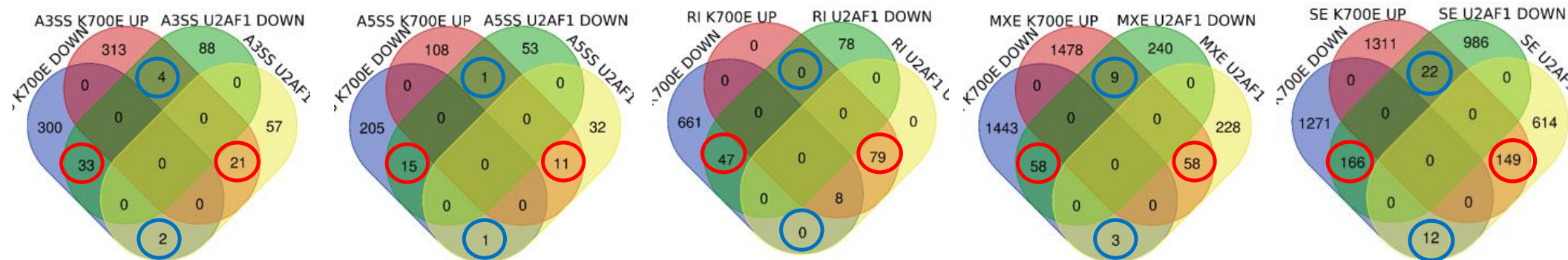


Overlap of genes/events undergoing AS regulation in SRSF2-P95 vs U2AF1 and SF3B1-K700E vs U2AF1 mutated patients

AS regulation in SRSF2-P95 vs U2AF1

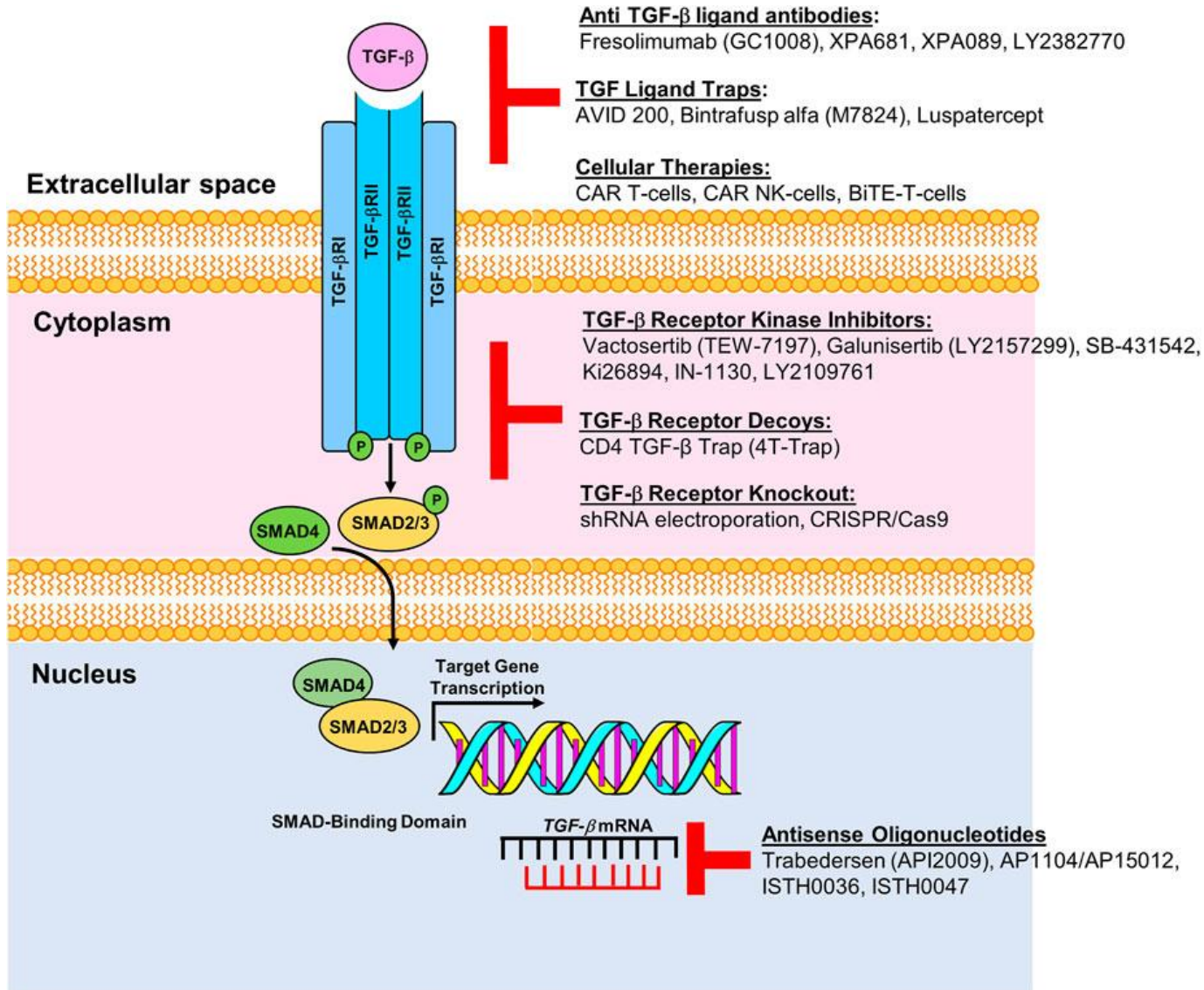


AS regulation in SF3B1-K700E vs U2AF1





Alternative Splicing in TGF-β Signaling Pathway



TGFBR2, transforming growth factor, beta receptor II

- The protein encoded by this gene is a transmembrane protein that has a protein kinase domain, forms a heterodimeric complex with TGF-beta receptor type-1, and binds TGF-beta
- This receptor/ligand complex phosphorylates proteins, which then enter the nucleus and regulate the transcription of genes related to cell proliferation, cell cycle arrest and tumorigenesis
- Deregulation of TGF-β pathway can be overcome by targeting the TGF-β receptors with ligand antibodies, ligand traps or by inhibiting TGF-β receptors using specific kinase inhibitors or by knocking out the TGF receptor genes with antisense oligonucleotides (e.g., AP11014 and AP15012)

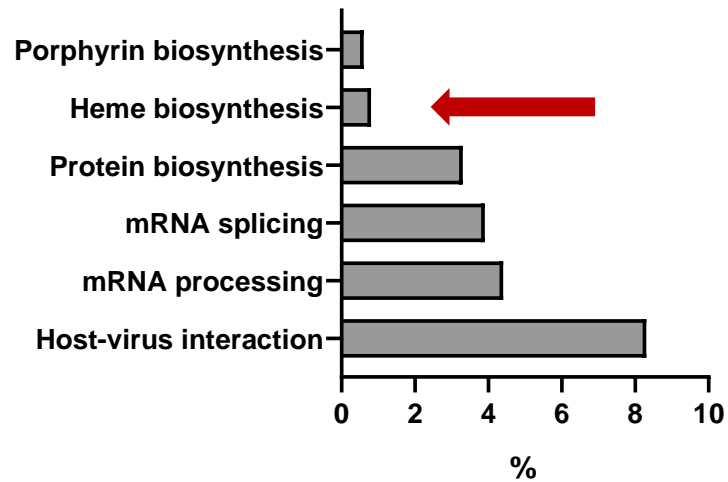


Functional annotation of AS events based on mutational status

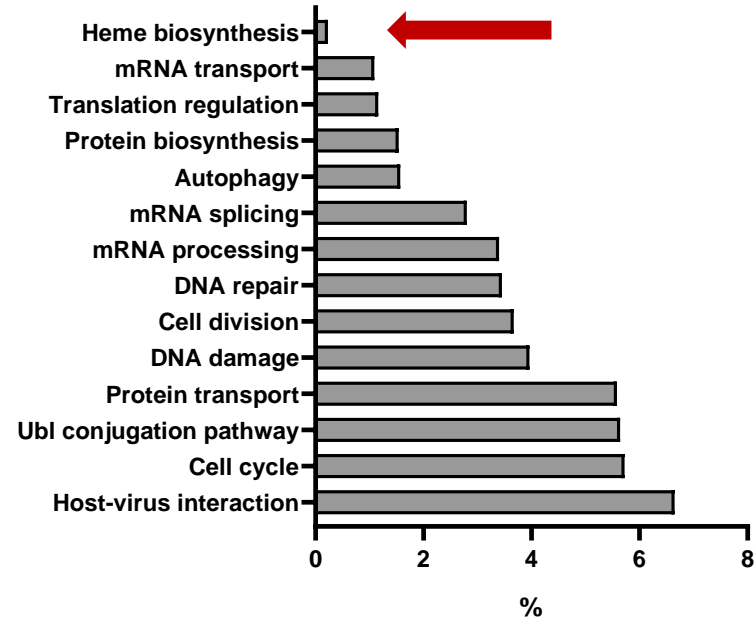


The heme biosynthesis pathway is altered in LR-MDS SF3B1 and SRSF2 mutated patients

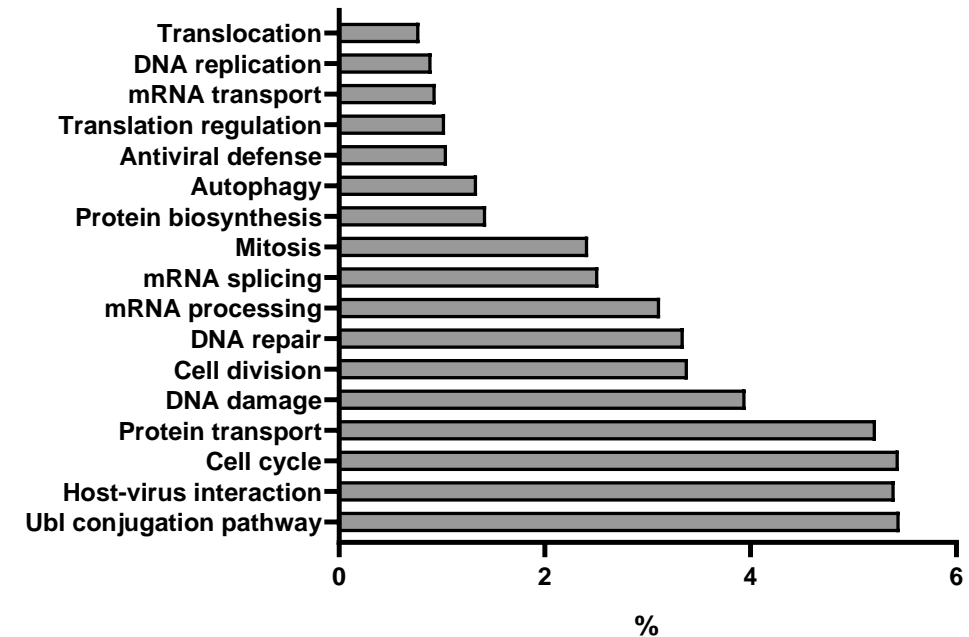
SF3B1-K700E



SRSF2- P95H/L/A

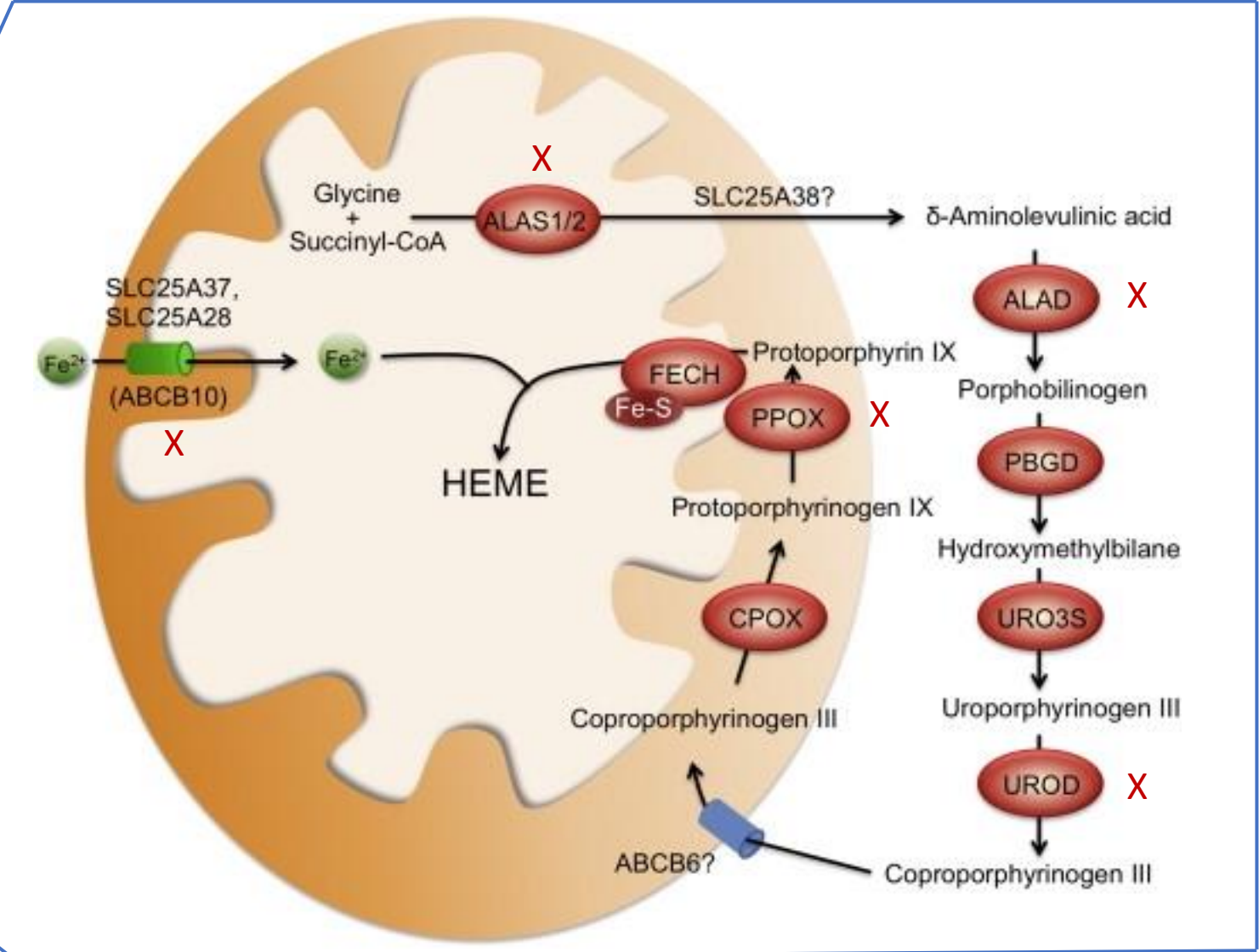
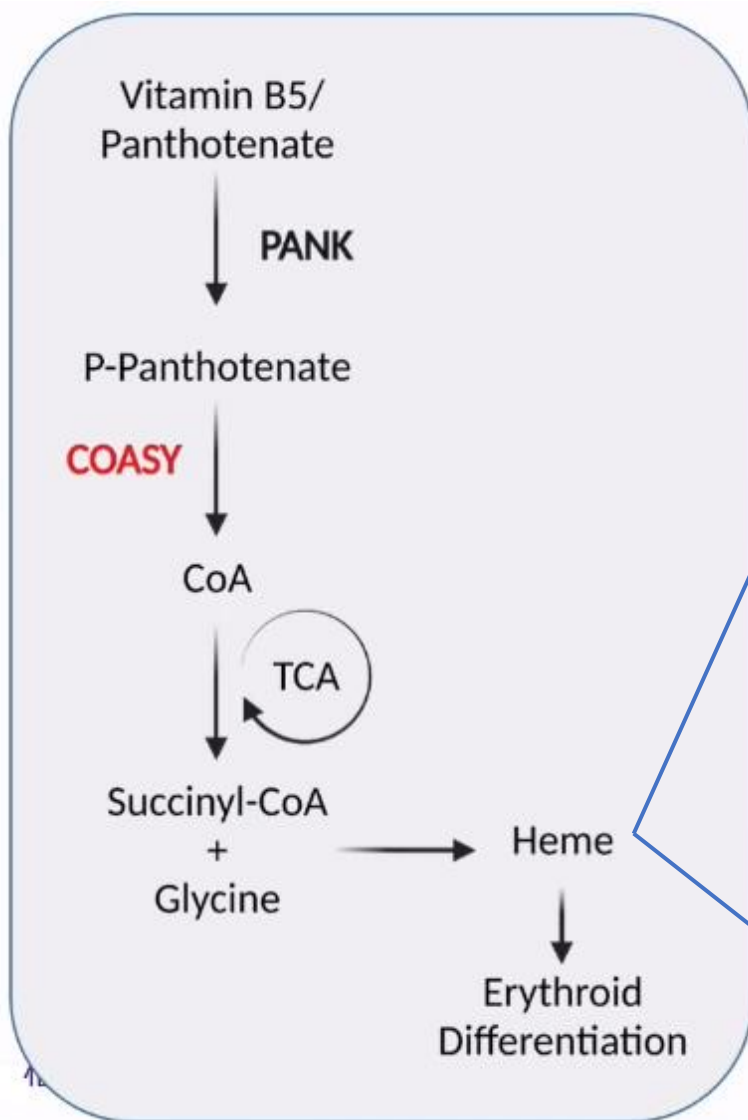


U2AF1-S34 and -Q157





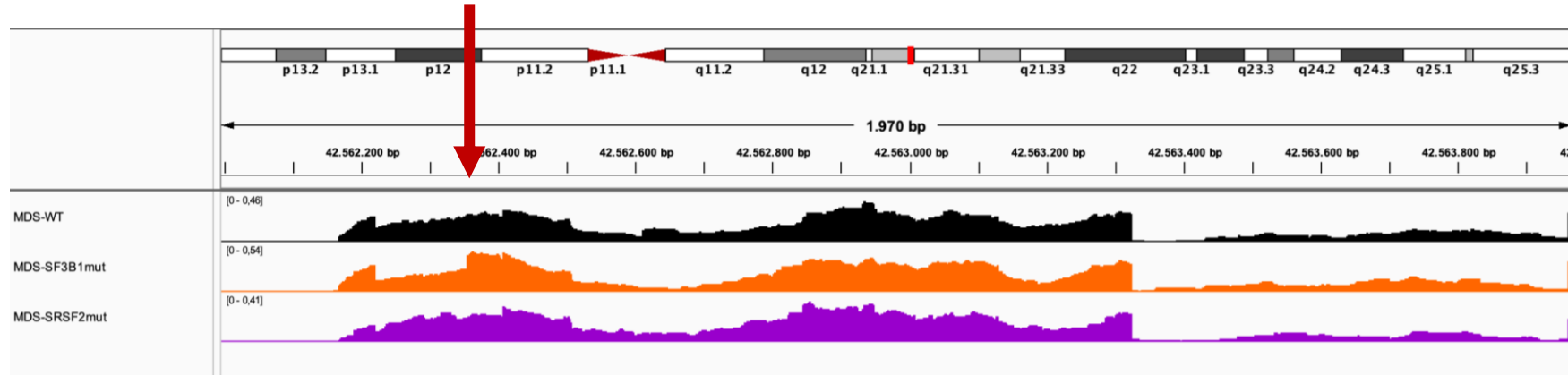
COASY deregulation impacts CoA synthesis and erythroid differentiation



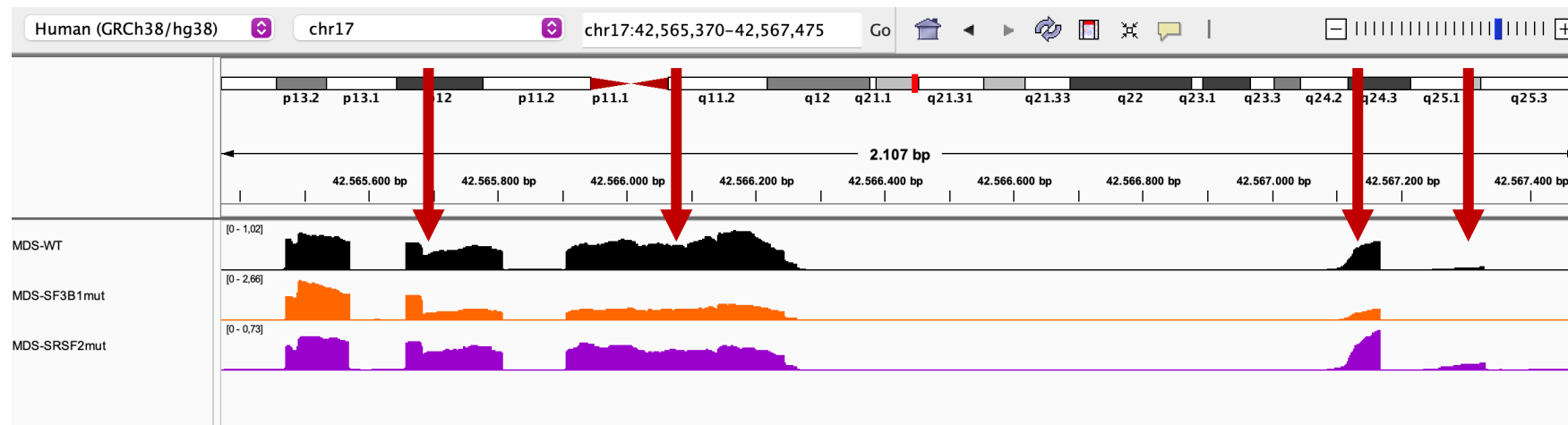


COASY: Alternative Splicing Events

Annotated event (PubMed)



Unannotated events





Conclusions



- RNA-Seq analysis showed a strong difference in gene expression profile between LR-MDS and non-hematological patients
- MDS with *SF3B1*^{K700E} showed a distinctive transcriptomic profile, while SRSF2 and U2AF1 mutated patients showed a more heterogeneous one
- In splicing factors mutated patients the alternative splicing events seem to be prominent compared to gene expression profile
- TGF- β pathway was identified as one of the biological pathways differently expressed in *SF3B1*^{K700E} patients, *suggesting its potential role* in the pathogenesis of MDS
- Recent findings suggest the involvement of COASY enzyme and genes belonging to HEME metabolism in erythroid differentiation
- Vitamin B5 and succinyl-CoA may improve ineffective erythropoiesis in *SF3B1* mutated MDS (Philippe *et al.*, 2023)

Grazie a tutti voi per l'attenzione

*Università di Roma Tor Vergata
Dipartimento di Biomedicina e Prevenzione*

Laboratorio di Oncoematologia

Prof.ssa Maria Teresa Voso
Dr Hajro Hajrullaj
Dr Antonio Cristiano
Dr.ssa Giorgia Silvestrini
Dr Angelo Onorato
Dr.ssa Elisa Galossi
Dr.ssa Giulia Falconi

Anatomia Umana

Prof.ssa Pamela Bielli
Dr Marco Pieraccioli
Dr.ssa Martina Valenzuela



*Department of Neuroscience,
Section of Human Anatomy,
Catholic University of the Sacred
Heath*

Prof Claudio Sette

MLL
Munich Leukemia Laboratory

Prof Torsten Haferlach
Dr Niroshan Nadarajah
Dr Stephan Hutter



8° WORKSHOP

In Ematologia Traslazionale
Della Società Italiana di Ematologia Sperimentale

